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Phylogeny and classification of the subfamily Rubioideae (Rubiaceae)

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Abstract. We performed phylogenetic analyses of the subfamily Rubioideae (Rubiaceae) based on three different pieces of chloroplast DNA, the protein coding rbcL gene, the spacer sequence between atpB and rbcL (atpB-rbcL), and the recently published (Andersson and Rova 1999) rps16 intron data. New rbcL sequences have been produced for 41 taxa and there are 52 new atpB-rbcL spacer sequences. All analyses gave similar results concerning the phylogeny, but they differ slightly in resolution and support for the various branches. The minor tribes Ophiorrhizeae, Urophylleae, Lasiantheae, and Coussareeae form a grade to the rest of the subfamily, which consists of two well-supported branches, the Psychotrieae alliance and the Spermacoceae alliance, including a majority of all genera and species. Based on the resulting phylogenies we present a revised classification of the Rubioideae. We accept 16 tribes of which 12 more or less correspond to earlier tribal circumscriptions: Anthospermeae, Argostemmateae, Craterisper-Gaertnereae, Morindeae, Paederieae, Psychotrieae, Schradereae, Spermacoceae, Rubieae, Theligoneae, and Urophylleae; two tribes have received new and very different circumscriptions: Ophiorrhizeae and Coussareeae; and two are new to science: Lasiantheae and Danaideae.

Key words: Rubiaceae, Rubioideae, Danaideae, Lasiantheae, chloroplast DNA, rbcL, atpB-rbcL spacer, rps16 intron, phylogeny, classification.

Progress in understanding of the subfamily Rubioideae of the Rubiaceae is relatively recent and includes many important contributions from many different scientists. Before the middle of the 20th century the "Rubioideae" taxa were dispersed in the two subfamilies Coffeoideae and Cinchonoideae, a classification of the Rubiaceae based on ovule number (Schumann 1891). Bremekamp (1952, 1954) and Verdcourt (1958) argued against this artificial division of the family and instead proposed that all Rubiaceae tribes with species containing raphides (calcium oxalate crystals) should be set aside as a new subfamily, Rubioideae. There are large similarities between the systems of these two authors. Verdcourt included 16 tribes (and the unplaced genus Hillia) in the subfamily Rubioideae, and Bremekamp (1966), who used a more narrow tribal circumscription, included 19 tribes and excluded some tribes to other subfamilies: Hillieae to Hillioideae, Ophiorrhizeae to Ophiorrhizoideae, Urophylleae and Pauridiantheae to Urophylloideae. Characters, except the raphides, that were used to distinguish Rubioideae were valvate aestivation (with few exceptions, Hamelia, Deppea, and Hillia – all these have later been shown to belong to Cinchonoideae; e.g. Bremer et al.

1995) and indumentum of septated (articulated) hairs. The next major change in the Rubioideae circumscription was presented by Robbrecht (1988). His Rubioideae with 16 tribes agreed largely with Bremekamp's but Robbrecht included Ophiorrhizeae and Theligoneae (shown to be related to Rubiaceae by Wunderlich 1971) and excluded Knoxieae Craterispermeae the subfamily and to Antirheoideae. three These classification schemes of Bremekamp, Verdcourt, and Robbrecht have been important sources of inspiration for later more focused studies of taxonomy and morphology. Comprehensive taxonomic studies of different Rubioideae tribes that should be mentioned are those by Puff and Robbrecht with co-authors (Puff 1982, 1988; Puff and Robbrecht 1989; Puff and Buchner 1998; Robbrecht 1982; Robbrecht et al. 1991), and Johansson (1987a, b, 1988, 1989, 1994). Other examples are chromosome investigations by Kiehn (e.g. 1986, 1995), morphological studies by Igersheim and co-authors (Igersheim 1992, Igersheim and Rohrhofer 1993, Igersheim et al. 1994), and pollen investigations by e.g. Robbrecht and coworkers (Robbrecht 1982, Jansen et al. 1996).

The classification systems as well as the taxonomic/morphological investigations have in their turn inspired phylogenetic investigations based on molecular data. In many of these, Rubioideae taxa have been included even if the focus has been on other groups or at other taxonomic levels. Several studies have aimed at resolving the whole family phylogeny (Bremer and Jansen 1991, Bremer et al. 1995, Bremer 1996a), others at specific tribes, e.g. Rubieae (Manen et al. 1994, Manen and Natali 1995, Manen and Natali 1996, Natali et al. 1996), and Psychotrieae (Nepokroeff et al. 1999). These studies are based on molecular markers designed to elucidate phylogenies at different taxonomic levels but the results are congruent to a great extent. The results in some parts agree with classification but in other parts they are totally different and there are several important conclusions concerning Rubioideae, e.g. that Hamelieae (Bremer and Jansen 1991) and Hillieae (Bremer et al. 1995, Natali et al. 1995) do not belong in the subfamily, that Theligoneae belong in the Rubioideae close to Rubieae (Bremer 1996a, Natali et al. 1996), and that Knoxieae belong in the Rubioideae and are not part of Antirheoideae (Bremer 1996a).

Molecular investigation has proved to be very informative for reconstruction of phylogeny but several tribes are yet to be represented in molecular investigations. Just before completion of this manuscript a detailed analysis of the Rubioideae based on the *rps*16 intron was published (Andersson and Rova 1999). That study confirms, in most parts, earlier molecular studies (Bremer 1996a, Natali et al. 1996) but as more taxa are included it also contributes a lot of new information.

The present study aims at presenting new phylogenetic analyses of the Rubioideae based on two different sequences of chloroplast DNA, the protein coding rbcL gene and the spacer sequence between atpB and rbcL (atpB-rbcL), a combined analysis of these data, and also an analysis of our data in combination with the recently published rps16 data, rbcL/atpB-rbcL/rps16 (also referred to as the 3-data matrix or analysis). As a result of the phylogenetic analyses we present a new classification of the subfamily Rubioideae.

Material and methods

The sampling strategy in the phylogenetic analyses was to cover as many of the described tribes of the subfamily Rubioideae as possible, and also to include several taxa from large and diverse tribes. We did not manage to find material from two of the twenty-eight tribes (Lathraeocarpeae and Perameae) and for economical and technical reasons we did not sequence all taxa for both rbcL and the spacer. All 151 genera included in the resulting classification are listed in the appendices 1 and 2.

The outgroups were chosen to represent one Gentianales member outside the Rubiaceae, *Gelsemium* (Gelsemiaceae; in the spacer and combined analyses), and members of other subfamilies of the Rubiaceae (cf. Bremer 1996a), *Gardenia* from the subfamily Ixoroideae, and *Cinchona* and

Luculia (in the spacer analysis also Coptosapelta and Exostema) from the subfamily Cinchonoideae. In the analysis of our data in combination with the rps16 data we used Gelsemium and Cinchona as outgroups.

Silica gel-dried or herbarium leaves were used in the DNA investigations. DNA was extracted. amplified, and sequenced according to Bremer et al. (1995) and Backlund et al. (2000) for rbcL, and according to Manen et al. (1994) for the atpB-rbcL spacer. Table 1 provides a list of taxa included in the phylogenetic analyses of rbcL and atpB-rbcL data, with EMBL/GenBank accession number of the corresponding sequences, and with voucher information for new sequenced taxa. New rbcL sequences have been produced for 41 taxa and there are 52 new atpB-rbcL spacer sequences. All other sequences used in these analyses have been published earlier by the first or second author alone or in collaboration with co-authors (Olmstead et al. 1993; Bremer et al. 1995; Bremer 1996a, b; Andreasen and Bremer 1996; Manen et al. 1994; Manen and Natali 1995, 1996). For references to the rps16 data see Andersson and Rova (1999).

The rbcL and atpB-rbcL spacer matrices comprise 106 and 63 taxa, respectively. The combined rbcL/atpB-rbcL matrix comprises 59 taxa. Most of the taxa used in the combined matrix are of the same species. However, Coprosma, Spermacoce, Manettia, Faramea, Coussarea, Praravinia, Hamelia and Gardenia are represented by two distinct species in the rbcL and in the atpB-rbcL matrix, respectively. In one analysis we included rps16 data from Anderson and Rova (1999) and performed a combined analysis. In this 3-data matrix we included 42 genera from the two studies. In several cases we had to compare different species from a genus, but in genera suspected to be paraphyletic, e.g., Psychotria and Hedyotis, we selected species that are supposed to be close and belong to the same monophyletic group.

The alignments of the *atpB-rbcL* spacer and the *rps*16 matrices were first done by the Clustal program (Thompson et al. 1994) followed by manual corrections. Some short regions could not be aligned and were excluded from the analyses. Homologous indel events were coded as presence/absence (1/0). When it was not possible to decide for homology, the event was coded with a questionmark. The aligned matrices are available on request or at http://www.unige.cjb.ch. The *rbcL* matrices

were without indels and the alignments were simple and without any alternatives.

The parsimony analyses were done using PAUP 3.1 (Swofford 1993). Only phylogenetically informative characters were included. The search method for the combined matrices was heuristic with 100 replications of RANDOM stepwise additions of sequences, the TBR branch swapping, and MULPARS options in effect. Support of the clades was calculated with bootstrap analyses with 1000 replicates and TBR branch swapping and MULPARS off.

Results

The *atpB-rbcL* spacer matrix (63 taxa) comprises 970 aligned positions, 453 of which are variable and 255 are potentially informative, 35 of which are indel characters. The *rbcL* matrix (106 taxa) comprises 1 402 sites, 560 of which are variable and 404 are potentially informative. The combined *rbcL/atpB-rbcL* matrix (59 taxa) comprises 2 364 sites, 899 of which are variable and 533 are potentially informative. The combined *rbcL/atpB-rbcL/rps*16 matrix (42 taxa) comprises 3 254 aligned positions, 1 269 of which are variable and 737 are potentially informative, of which 57 are indel characters.

The atpB-rbcL spacer analysis (Fig. 1). The bootstrap analysis of the 63 spacer sequences (52 from Rubioideae and eleven outgroup taxa) resulted in a tree illustrated in a simplified form showing mainly the relationships of the tribes (Fig. 1).

In the atpB-rbcL spacer tree, based on 255 phylogenetically informative characters, the mean bootstrap value for the nodes is 55%, and 35% of the nodes have bootstrap values $\geq 75\%$.

In agreement with earlier studies (Bremer et al. 1995, Natali et al. 1995, Andersson and Rova 1999) the tribes Hamelieae and Hillieae do not belong in the subfamily Rubioideae but instead in the Cinchonoideae. *Virectaria* (former Virectarieae) is close to *Gardenia* of the Ixoroideae (as in Bremer and Thulin 1998).

The subfamily Rubioideae is well-supported with a bootstrap value of 100%. The base of the

Table 1. List of taxa sequenced for *rbc*L and *atp*B-*rbc*L. Earlier unpublished sequences are indicated with a * and voucher information is given. All other taxa have been published earlier by the first or second author alone or in collaboration with co-authors (see Material and Methods). Names of herbaria are abbreviated according to Holmgren et al., 1990. All *rps*16 sequences analysed in this paper are from Andersson and Rova 1999

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/ voucher information
Agathisanthemum bojeri (Hiern.) Klotzsch	Z68787		
Amaracarpus Blume sp. Amphiasma luzuloides (K. Schum.) Bremek.	AJ002176 AJ288594*		Tanzania, Herb. material, Iversen et al. 87694 (UPS)
Amphidasya ambigua (Standley) Standley	Y11844		` ,
Anthospermum herbaceum L.f.	X83623	AJ234028*	Tanzania, Bremer 3093 (UPS)
Appunia guatemalensis Donn. Smith	AJ288593*	AJ234009*	North America, Herb. material, Martinez 13581 (G
Arcytophyllum aristaum Standley	AJ288595*		Ecuador, Bremer et al. 3371 (UPS)
Argostemma hookeri King Asperula laevigata L.	Z68788 X81092	AJ234032*	Malaysia, Wanntorp s.n. (S)
Batopedina pulvinellata E. Robbrecht	AJ288596*		Zaire, Herb. material, Malaisse 7695 (UPS)
Bouvardia glaberrima Engelm.	X83626		
Bouvardia glaberrima Engelm.		X76478	
Carpacoce Sond. sp.	AJ288597*		South Africa, Bremer 3708 (UPS)
Carphalea glaucescens (Hiern.) Verdc.	Z68789		, ,
Chasalia parviflora Benth.	Z68790		
Chazaliella abrupta (Hiern) E. Petit & Verd.	Z68791		
Cinchona pubescens Vahl	X83630		
Cinchona pubescens Vahl		AJ233990*	Jamaica, McDowell. 4613 (DUKE)
Coccocypselum hirsutum (Bartling ex DC.) L.O. Williams	X87145		
Coccocypselum P. Br. sp.		X811678	
Coelospermum balansanum Baill.	AJ288598*	AJ234010*	New Caledonia, Herb. material, Veillon 3765 (G)
Commitheca liebrechtsiana (De Wild & Th. Dur.) Bremek.		AJ233999*	Gabon, Herb. material, Vilks 478 (ME)
Conostomium quadrangulare (Rendle) Cufod.	Z68792		

Table 1. (Continued)

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/voucher information
Coprosma ernodeoides A. Gray		AJ234029*	Hawaii, Kiehn 910114 (WU)
Coprosma pumila Hook. f.	X87146		
Coptosapelta diffusa (Champ. ex Beneth.) van Steenis	A0/140	AJ233987*	China, Herb. material, Bartholomew et al. 1188 (ME)
Coussarea contracta Beneth. & Hook. f.		AJ234007*	Paraguay, Zardini and Velasquez 9774 (G)
Coussarea macrophylla Muell. Arg.	Y11847		, 0.004,002 5, , , (0)
Craterispermum brachynematum Hiern	AJ288629*	AJ234011*	Zaire, Herb. material, Lejoly 2707 (ME)
Cruciata glabra (L.) Ehrend.	X81097		
Cruckshanksia hymenodon Hook. & Arn.	AJ288599*		Chile, Herb. material, Rodriguez 10 (K)
Cruckshanksia hymenodon Hook. & Arn.		AJ234004*	Chile, Herb. material, Billiet and Jardin 5570 (ME)
Damnacanthus indicus Gaertn. f.	Z68793	AJ234015*	Cult., Missouri Bot. Gard. Bremer 3107 (UPS)
Danais xanthorrhoea (K. Schum.) Bremek.	Z68794	AJ234019*	Tanzania, Bremer 3079 (UPS)
Declieuxia fruticosa Kuntze Didymaea alsinoides (Cham. & Schlecht.) Standley	AJ002177 Z68795		
Didymaea alsinoides (Cham. & Schlecht.) Standley		AJ234036*	Costa Rica, Kiehn sn (WU)
Diodia sarmentosa SW. Ernodea litoralis SW. Exostema caribaeum	AJ288600* AJ288601* X83636	AJ234025*	Puerto Rico, Taylor 11749 (MO) Cuba, Kiehn sn (WU)
(Jacq.) Roem. & Schult. Exostema caribaeum (Jacq.) Roem. & Schult.		AJ233991*	Cuba, Kiehn 910526 (WU)
Faramea multiflora A. Rich. Faramea porophylla	Z68796	AJ234008*	Paraguay, Herb. material,
Muell. Arg. Gaertnera Retz. sp.		AJ234012*	Zardini 8630 (G) Madagascar, Herb. material, Malcomber 999 (BR, G, MO, TAN, WAG)
Gaertnera Retz. sp.	Z68797		•
Gaillonia yemenensis Thulin	AJ288630*		Yemen, Thulin et al. 9365 (UPS)
Galium album Mill.	X81090		
Galopina circaeoides Thunb.	AJ288602*		South Africa, Bremer 3797 (UPS)

Table 1. (Continued)

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/voucher information
Gardenia taitensis DC.		AJ233988*	Cult., Geneva Bot. Gard., Natali and Manen 007 (G)
Gardenia thunbergia Linn. f.	X83637		1 www was 17 was 507 (3)
Gelsemium sempervirens Ait.	L14397	AJ233985*	
Geophila repens (L.) I.M. Johnston	Z68798	AJ234017*	Cult., Uppsala Bot. Gard. Bremer 3130 (UPS)
Gynochthodes coriacea Blume	AJ288603*		Cult., Bogor Bot. Gard., Ridsdale XVII.C.103 (L)
Hamelia cuprea Griseb.	X83641		
Hamelia papillosa Urb.		AJ233992*	Jamaica, McDowell. 4600 (DUKE)
Hedyotis caerulea Wight & Arn.	AJ288604*		Cult., Uppsala. Bot. Gard. Bremer s.n. (UPS)
Hedyotis fruticosa Linn.	Z68799	AJ234026*	,
Hedyotis littoralis (Hillebr.) Fosberg "Kadua"	AJ288605*	AJ234027*	Cult., Vienna Bot. Gard., Kiehn & Luegmayr 920823 (WU)
Hedyotis nigricans (Lam.) Fosberg	AJ288606*		USA, Miller et al., 8232 (MO)
Hemidiodia ocimifolia Schum.	AJ288607*		Ecuador, Bremer et al., 3340 (MO, QCA, QCNE, UPS)
Hillia triflora (Oerst.) C.M. Taylor	X83642	AJ233993*	Cult., Univ. of Colorodo, Bremer 3101 (UPS)
Hillia valeris Standley	AJ288608*	X81683	Costa Rica, Kiehn 880331 (WU)
Hoffmannia refulgens Hemsl. x ghiesbreghtii Hemsl.	X83644		
Hydnophytum formicarum Jack	X83645		
Hydnophytum formicarum Jack		X76480	
Hymenocoleus hirsutus (Benth.) Robbrecht	AJ002178		
Knoxia platycarpa Arn.	AJ288631*		Sri Lanka, Herb. material, Lundqvist 11302 (UPS)
Kohautia caespitosa Schnizl.	Z68800		- , ,
Lasianthus pedunculatus E.A. Bruce	Z68802	AJ234003*	Tanzanaia, Andreasen 71 (UPS)
Lelya prostrata (Good) W.H. Lewis	AJ288609*		Malawi, Herb. material, Thomson & Rawlins 5482 (K)
<i>Lerchea bracteata</i> Valeton	AJ288610*	AJ233997*	Sumatra, Herb. material, Axelius 343 (S)
Luculia grandifolia Ghose	X83648	AJ233986*	Cult., Stockholm Univ., Bremer 2713 (S)
Manettia bicolor Paxt.	Z68803		

Table 1. (Continued)

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/ voucher information
Manettia cordifolia Mart.		AJ234022*	Argentina, Herb. material, Novara 9814 (G)
Mapouria cf. umbrosa Muell. Arg.	Z68804		
Maschalocorymbus corymbosus (Blume) Bremek.	AJ288611*		Sabah, Ridsdale 2471 (L)
Mitchella repens Linn.	Z68805		
Mitchella Linn. sp.		AJ234016*	Japan, Ehrendorfer 930905-1601 (WU)
Mitracarpum villosum Cham. & Schlecht.	AJ288632*		Sri Lanka, Herb. material, Fagerlind 810 (S)
Morinda citrifolia Linn.	X83651	AJ234013*	Cult., Uppsala Bot. Gard., Bremer 3106 (UPS)
Mycetia malayana Craib	Z68806	AJ234033*	Cult., Univ. of Aarhus, Larsen et al., 42486 (AAU, UPS)
Myrmecodia platyrea Becc.	X87147		,
Nertera granadensis (L.F.) Druce	X83654		
Neurocalyx zeylanicus Hook.	Z68807	AJ233995*	Sri Lanka, Bremer 937 (PDA, S, US)
Oldenlandia cf. corymbosa Linn.	X83655		(, -,,
Oldenlandia goreensis Summerhayes	Z68808		
Opercularia vaginata Labill.		AJ234030*	Australia, Herb. material, Weber 9157 (G)
Opercularia vaginata Labill.	Z68809		(-)
Ophiorrhiza mungos Linn.	X83656		Cult., Meisse Bot. Gard. Robbrecht s.n. (UPS)
Ophiorrhiza Linn. sp.		X81677	Sumatra, Frimmel s.n. (WU)
Oreopolus glacialis	AJ288612*		Argentina, Swenson 328 (UPS)
(Poepp. & Endl.) Ricardi			
Otiophora cupheoidea N.E. Br.	AJ288613*		South Africa, Bremer 3805 (UPS)
Otomeria oculata S. Moore	AJ288614*		Ethiopia, Herb. material, Puff 821222-2/1 (K)
Paederia foetida Linn.		AJ234006*	Japan, Ehrendorfer 930830-0801 (WU)
Palicourea Aubl. sp.	Z68810		• •
Parapentas silvatica (K. Schum.) Bremek.	X83657	AJ234021*	Tanzania, Bremer 3091 (UPS)
Paratriainia xerophila Bremek.	AJ288633*		Madagascar, Herb. material, Croat 30548 (MO)
Pauridiantha paucinervis (Hiern) Bremek.	Z68811	AJ233998*	Tanzania, Bremer 3090 (UPS)

Table 1. (Continued)

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/ voucher information
Pentanisia longituba	Z68812		
Oliver		AJ234022*	South Africa, Herb.
Pentanisia microphylla Chior.		AJ254022	material, Moll 610 (G)
Pentanopsis fragrans Rendle	Z68813		,, (-)
Pentas lanceolata (Forsk.) Defl.	X83659		Cult., Univ. of Connecticut, Bremer 2702 (S)
Pentas lanceolata (Forsk.) Defl.		X76479	Geneva Bot. Gard., Natali and Manen 004 (G)
Pentodon pentandrus Vatke	X83660	AJ234024*	Tanzania, Bremer 3082 (UPS)
Phuopsis stylosa Beneth. & Hook. f.	X81103		
Phyllis nobla Linn.	Z68814		
Phyllis nobla Linn.		AJ234031*	Canary Islands, Herb. material, Charpin and Rodriguez 18576 (G)
Phylohydrax carnosa (Hochst.) C. Puff	AJ288615*		South Africa, Bremer 3783 (UPS)
Placopoda virgata Bolf. f.	Z68815		
Plocama pendula Ait.	Z68816	AJ234035*	Canary Islands, Andreasen 1 (UPS)
Praravinia densiflora Korth. Praravinia suberosa	AJ288616*	AJ234000*	Borneo, Clemens 33777 (G) Sabah, no voucher, Ridsdale
(Merill) Bremek. Pravinaria leucocarpa Bremek.	AJ288617*	AJ234001*	Borneo, Herb. material, Collenette 21654 (G)
Prismatomeris labordei (Leveille) Merill apud Rehder		AJ234005*	China, Herb. material, Bartholomew et al. 2118 (ME
Prismatomeris beccarianum (Baillon) J.T. Johansson	AJ288618*		Sabah, Herb. material, Ridsdale 2461 (L)
Psychotria kirkii Hiern (bacteriophila) Yalet.	X83663		
Psychotria kirkii Hiern (bacteriophila) Yalet.		X76481	
Psychotria peteri Verdcourt	Z68817		
Psychotria poeppigiana Muell. Arg.	Z68818	AJ234018*	Ecuador, Bremer et al. 3030 (MO, QCA, QCNE, UPS)
Psychotria Linn. sp. "cephaelis"	AJ002188		
Psyllocarpus laricoides Mart.	AJ288619*		Brasil, Herb. material., Andersson et al. 355750 (UPS)

Table 1. (Continued)

	Accession EMBL/ GenBank rbcL	Accession EMBL/ GenBank atpB-rbcL	Source/voucher information
Putoria calabrica	AJ288620*		Herb. material, Jonsell
(L.) DC.			4216 (UPS)
Putoria calabrica		X81672	Greece, Ehrendorfer
(L.) DC.			930409-2301 (WU)
Relbunium hypocarpium Hemsl.	AJ288621*		Cult., Meise Bot. Gard., Billiet 3788 (BR)
Richardia pilosa Ruiz. & Pav.	Z68820		
Rubia tinctorum Linn.	X83666		
Rubia tinctorum Linn.		X76465	
Rudgea sessiliflora Standley	AJ002186	11,0,00	
Rutidea orientalis D.M. Bridson	Z68862		
Schenckia blumenaviensis K. Schum.	AJ288622*		Cult., Copenhagen Bot. Gard., Ryding 2359 (C)
Schenckia blumenaviensis K. Schum.		AJ233994*	Brazil, Kiehn sn (WU)
Schradera subandina Krouse	Y11859	AJ234014*	Ecuador, Clark & Watt 783 (MO, QCNE, UPS)
Serissa fetida Lam.	Z68822	AJ234034*	Cult., Univ. of Connecticut, Bremer 2717 (UPS)
Sherardia arvensis Linn.	X81106		Stemer 2/17 (CFS)
Spermacoce assurgens Ruiz & Pav.		X81679	
Spermacoce hispida Linn.	AJ288623*		Sri Lanka, Herb. material, Wanntorp et al. 2667 (S)
Spermacoce laevis Roxb.	Z68823		• • • • • • • • • • • • • • • • • • • •
Spermadictyon suaveolens Roxb.	Z68824		
Synaptantha tillaeacea (F. Muell.) J.D. Hook.	AJ288624*		Australia, Herb. material, Lazarides & Palmer 272 (K)
Theligonum cynocrambe Huth		X81680	Greece, Ehrendorfer 930416-6201 (WU)
Theligonum cynocrambe Huth	X83668		
Triainolepis hildebrandtii Vatke	AJ288625*	AJ234020*	Kenya, Herb. material, Bally 13258 (G)
Trichostachys Hook f. in Beneth. & Hook. f. sp.	AJ288626*		Cameroun, Sonké 1725 (UPS)
Urophyllum ellipticum Thw.	AJ288627*	AJ234002*	Herb. material, Lundqvist 11085 (UPS)
Valantia muralis L.	X81107		
Virectaria major K. Schum.	Y11861	AJ233989*	Herb. material, Reekmans 10916 (UPS)
Xanthophytum capitellatum Ridley	AJ288628*	AJ233996*	Sabah, Ridsdale 2473 (L)

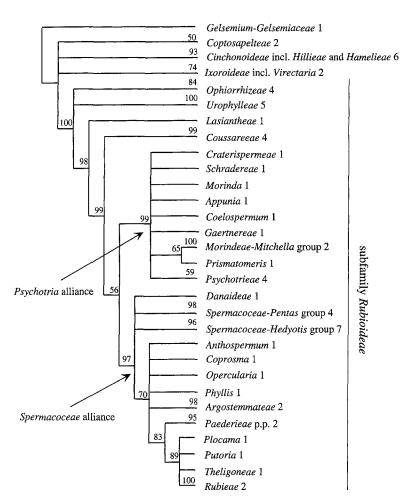


Fig. 1. Simplified phylogenetic tree illustrating the relationships of Rubioideae based on *atp*B-*rbc*L sequences (analysis includes 52 ingroup taxa). Numbers above nodes indicate bootstrap values (≥50%) from 1000 replicates with TBR branch swapping, all bootstrap values ≥50% are indicated. Numbers behind names represent number of investigated taxa on this branch

Rubioideae forms a trichotomy. One branch is the Ophiorrhizeae (bootstrap 84%), another is the Urophylleae (bootstrap 100%), and the third branch includes the remaining part of the subfamily (bootstrap 98%). In this large clade, Lasianthus (Lasiantheae) is the first taxon to branch off followed by the clade Coussareeae (bootstrap 99%, including Coussarea, Faramea, Cruckshanksia, former Hedyotideae/Cruckshanksieae and Coccocypselum). The sister group to Coussareeae is not well-supported (56%). However, within this clade there are two well-supported subclades. One of them here called the Psychotrieae alliance (bootstrap 99%) contains Pychotrieae, Craterispermeae, Gaertnereae, Schradereae, and Morindeae (not supported as monophyletic). The other branch (bootstrap 97%) is here called the Spermacoceae alliance and contains Spermacoceae (not supported as monophyletic), Anthospermeae (not supported as monophyletic), Argostemmateae, Danaideae, Paederieae (in this analysis paraphyletic), Rubieae, and Theligoneae.

The *rbc*L analysis (Fig. 2). The bootstrap analysis of the 106 *rbc*L sequences resulted in a tree illustrated in a simplified form showing mainly the relationships of the tribes (Fig. 2).

The topology of this *rbcL* tree (of 102 Rubioideae taxa and four outgroup taxa) is similar to the spacer tree (Fig. 1) but the support values are slightly higher. In the *rbcL* tree, based

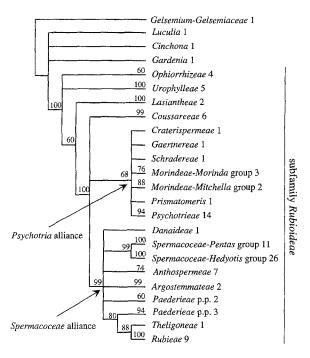


Fig. 2. Simplified phylogenetic tree illustrating the relationships of Rubioideae based on *rbc*L sequences (analysis includes 102 ingroup taxa). Numbers above nodes indicate bootstrap values (≥50%) from 1000 replicates with TBR branch swapping, all bootstrap values ≥50% are indicated. Numbers behind names represent number of investigated taxa on this branch

on 404 phylogenetically informative characters, the mean bootstrap value for the nodes is 60%, and 38% of the nodes have bootstrap values \geq 75%. All tribes that are supported in the spacer tree are also supported in the rbcL tree. Further, in the rbcL tree there is also support (74%) for a monophyletic Anthospermeae and a monophyletic Spermacoceae (99%), which are not supported in the spacer tree.

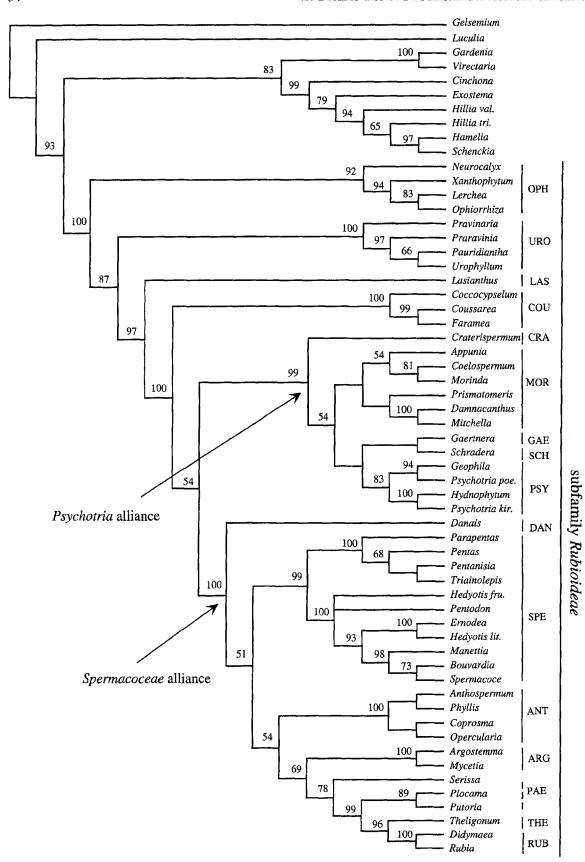
The combined *rbcL/atpB-rbcL* analysis (Fig. 3). The analysis of 59 taxa (49 Rubioideae taxa and ten outgroup taxa) from a combined matrix of *rbcL* and the spacer sequences resulted in two trees 1 787 steps long with a consistency index of 0.427 (excluding uninformative characters) and a retention index of 0.713. The strict consensus tree with bootstrap values indicated is presented in Fig. 3.

The analysis of the combined data produced trees very similar to the topology of the

spacer and rbcL trees. The main differences are the better resolution within the trees and much higher support values for many branches. In this combined tree, based on 533 phylogenetically informative characters, the mean bootstrap value for the nodes is 81%, and 68% of the nodes have bootstrap values $\geq 75\%$.

The Rubioideae clade is supported by 100%. The most basal clade to branch off within the Rubioideae is the Ophiorrhizeae (bootstrap 92%; including Ophiorrhiza, Lerchea, Neurocalyx, and Xantophytum), separated (87%) from the following Urophylleae 100%; including Urophyllum, (bootstrap Pravinaria. Praravinia, and Pauridiantha). High bootstrap values (97%) support the isolation of Lasianthus from the basal clades and the remaining taxa (100%). The sister group of Lasianthus comprises three main branches. The relationships between these are not supported (<50%). The three branches are the Coussareeae (bootstrap 100%, including here also Coccocypselum), the Psychotrieae alliance (bootstrap 99%), and the Spermacoceae alliance (bootstrap 100%). Inside the Psychotrieae alliance clade, the tribe Craterispermeae is sister to the remaining part (bootstrap 54%), which comprises Gaertnereae, Morindeae, Schradereae, and Psychotrieae. The relationships between these tribes are not supported (<50), and of these tribes only two are represented by several taxa of which only tribe Psychotrieae has high support (83%).

Inside the Spermacoceae alliance clade, the position of Danaideae has low support (bootstrap 51%). The remaining taxa are included in two clades, one is the Spermacoceae (bootstrap 99%) which are clearly divided in two: one subclade (bootstrap 100%) containing the *Pentas* group and former members of Knoxieae and Triainolepideae, and another subclade (bootstrap 100%) containing e.g. the *Spermacoce* and the *Oldenlandia* groups. The other main branch of the Spermacoceae alliance is not well-supported (54%) but includes the relatively well-supported clades of Anthospermeae (bootstrap 100%), Argostemmateae (bootstrap 100%), and Paederieae, Theligo-



neae, and Rubieae (bootstrap 78%). Paederieae are paraphyletic.

The combination of our rbcL/atpB-rbcL data with rps16 data (Fig. 4). The analysis of 42 taxa (38 Rubioideae taxa and four outgroup taxa) from a combined matrix of data from the rbcL, the atpB-rbcL spacer and the rps16 intron (Anderson and Rova 1999) resulted in 24 trees 2 096 steps long with a consistency index of 0.503 (excluding uninformative characters) and a retention index of 0.682. The strict consensus tree with bootstrap values indicated is presented in Fig. 4.

The topology of the consensus tree is almost identical or congruent with the *rbcL/atpB-rbcL* tree but the support value for each branch is higher, as could be expected since more characters are included. For example, the separation between the Coussareeae and Psychotrieae and Spermacoceae alliances are better supported (70% compared to <50%), and the Anthospermeae, Argostemmateae, Paederieae, Theligoneae, and Rubieae branch has higher support (91% compared to 54%). This 3-data tree, based on 737 phylogenetically informative characters, has a mean bootstrap value for the nodes of 81%, and 72% of the nodes have bootstrap values ≥75%.

Two minor differences between this tree and the combined rbcL/atpB-rbcL tree are that the trichotomy of Ophiorrhizeae, Urophylleae and the rest of the subfamily is unresolved, and that the relationships within the Psychotrieae are less resolved, but these relationship are not supported in the rbcL/atpB-rbcL (<50%) tree.

Classification. As a result of the phylogenetic analyses of molecular data in combination with morphological characters and comprehensive information from literature we here present a revised classification of the subfamily Rubioideae (below and Table 2). We have only listed

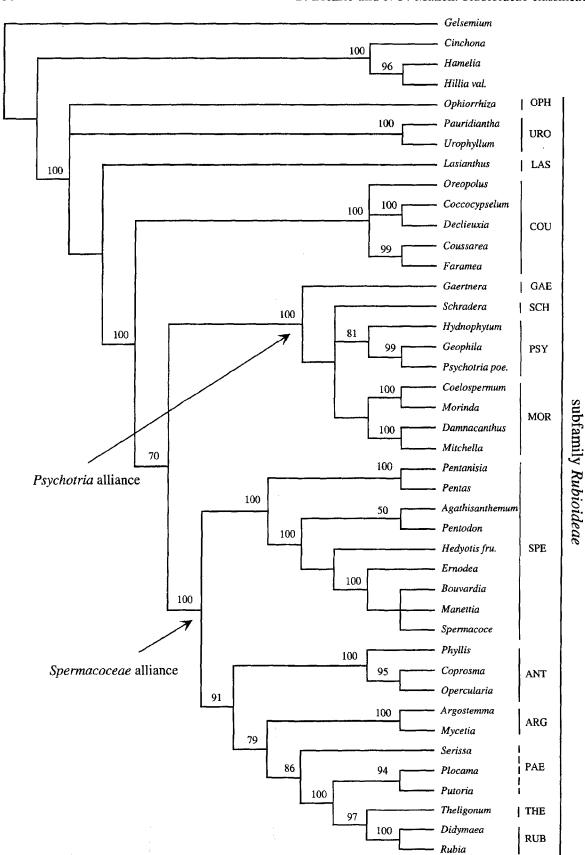
those genera that we have sequenced and genera with a close relationship to sequenced genera as judged from morphology.

Subfamily Rubioideae Verdcourt, Bull. Jard. Bot. État Brux. 28: 280 (1958)

- Cephaelidoideae Rafinesque, Ann. Gén.
 Sci. Phys. Brux. 6: 86 (1820)
- Richardioideae Rafinesque, Ann. Gén.
 Sci. Phys. Brux. 6: 84 (1820)
- Spermacocoideae Chevall., Fl. Gén. Env. Paris 2: 605 (1827) as "Spermacocceae"
- Anthospermoideae Kostel., Allg. Med.-Pharm. Fl. 2: 537 (1833) as "Anthospermeae"
- Hedyotidoideae Kostel., Allg. Med.-Pharm. Fl. 2: 538, 569 (1833) as "Hedyotideae"
- Opercularioideae Kostel., Allg. Med.-Pharm. Fl. 2: 537 (1833) as "Opercularieae"
- Paederioideae Kostel., Allg. Med.-Pharm. Fl. 2: 537, 564 (1833) as "Paederieae"
- Gaertneroideae Arn., Pug. Pl. Ind. Or.: 351, 352 (1836) as "Gaertnerieae"
- Coccocypseloideae Burmeist., Handb.
 Naturgesch., 286 (1837) as "Coccocypseleae"
- Manettioideae Burmeist., Handb. Naturyesch., 286 (1837) as "Manettieae"
- Aparinoideae Pfeiff., Nomencl. Bot. 1(1): 320 (1872)
- Pomazotoideae Bremekamp ex S. Darwin, Taxon 25: 605 (1976)

Shrubs, herbs, or less common trees. Raphides present. Heterostylous flowers common. Stipules entire, bifid or often fimbriate. Corolla aestivation always valvate. Ovary 1-12-locular, most often 2-locular either with many or with single ovules in each locule, more rarely two or few in each locule. Fruits dry or fleshy, dehiscent or indehiscent.

Fig. 3. Strict consensus tree of two equally parsimonious trees of Rubioideae based on rbcL and atpB-rbcL sequences, after a heuristic search with 100 replicates and TBR branch swapping. Numbers above nodes indicate bootstrap values ($\geq 50\%$) of 1000 replicates with TBR branch swapping, all bootstrap values $\geq 50\%$ are indicated. Vertical bars and corresponding letters (first three letters of the tribes cf. Table 2) represent the tribal classification according to Bremer and Manen



Tribes included (here investigated): Anthospermeae, Argostemmateae, Coussareeae, Craterispermeae, Danaideae, Gaertnereae, Lasiantheae, Morindeae, Ophiorrhizeae, Paederieae, Psychotrieae, Rubieae, Schradereae, Spermacoceae, Theligoneae, Urophylleae.

Uncertain status (not investigated): Lathraeocarpeae, Perameae.

Tribe Ophiorrhizeae Bremekamp ex Verdcourt, Bull. Jard. Bot. État Brux. 28: 281 (1958)

Pomazoteae Bremekamp ex S. Darwin,
 Taxon 25: 605 (1976)

Useful recent studies: Darwin (1976), Bremer (1979), Axelius (1987, 1990)

Herbs or subshrubs to small trees. Stipules entire, bifid to fimbriate. Flowers homo- or heterostylous, *Neurocalyx* with adnate anthers. Ovary 2-locular, with many ovules. Fruits dry, dehiscent (flattened capsules in *Ophiorrhiza*) or indehiscent with thin to thick pericarps, with many small (dust) seeds. Pollen 3-colporate. Chromosome basic number x = 11 (12) with 2-or 4-ploidy level (Kiehn 1995).

Genera included (here investigated): Ophiorrhiza, Neurocalyx, Lerchea, Xanthophytum.

Inclusion based on morphology: Coptophyllum (incl. Pomazota), Spiradiclis, (and included in Xanthophytum: Paedicalyx and Xanthophytopsis).

Tribe Urophylleae Bremekamp ex Verdcourt, Bull. Jard. Bot. État Brux. 28: 281 (1958)

- Pauridiantheae Bremekamp ex S. Darwin, Taxon 25: 605 (1976)

Shrubs, subshrubs, or small trees. Stipules entire or fringed. Ovary 2-pluri-locular. Fruits fleshy with many seeds. Flowers unisexual or hermaphroditic, homo- or heterostylous. Pollen 3-colporate. Chromosome basic number x = 9 with 2- or 6-ploidy level (Kiehn 1995).

Genera included (here investigated): Urophyllum, Amphidasya, Commitheca, Maschalocorymus, Praravinia, Pravinaria, Pauridiantha.

Urophyllum and Pauridiantha both have the rare basic number x=9 and they also have bimodal karyotypes (Kiehn: pers. comm.). In Andersson and Rova (1999) Raritebe belongs to the Urophylleae clade. Raritebe was earlier (Robbrecht 1993) placed in the Isertieae (subfamily Cinchonoideae) and reported not to contain raphides. We have not seen any material of the genus but if the position is correct it is the first report of a genus without raphides in Rubioideae.

Tribe Lasiantheae B. Bremer and Manen, trib. nov.

Dressleriopsideae Dwyer, Ann. Missouri.
 Bot. Gard. 67:11 (1980) - not validly published
 Type genus: Lasianthus

Frutices, arbores parvae vel suffrutices. Stipulae integrae saepe triangulares. Flores interdum heterostyli. Ovaria 2- vel 4-12-locularia, ovulo in quoque loculo singulari. Fructus drupacei, nec compositi nec aggregati, saepe cyanei ad ateri pyrenis 2 vel 4-12. – Differt a tribus affinibus fructibus cyaneis vel atris, pyrenis 4-12 (*Trichostachys* 2) et inflorescentiis saepe axillaribus sessilibus.

Shrubs, small trees, or subshrubs. Stipules entire, often triangular. Flowers sometimes heteostylous. Ovary 2- (*Trichostachys*) or 4-12-locular, with a single ovule in each locule, erect from the base. Fruits drupaceous, not compound or aggregated, often blue to black, with 2 or 4-12 pyrenes. Chromosome basic number x = 11 with 4- (Bir et al. 1984) or 22-ploidy level (Kiehn pers. comm.).

Genera included (here investigated): Lasianthus, Trichostachys.

Fig. 4. Strict consensus tree of 24 equally parsimonious trees of Rubioideae based on rbcL, atpB-rbcL, and rps16 sequences, after a heuristic search with 100 replicates and TBR branch swapping. Numbers above nodes indicate bootstrap values ($\geq 50\%$) of 1 000 replicates with TBR branch swapping, all bootstrap values $\geq 50\%$ are indicated. Vertical bars and corresponding letters (first three letters of the tribes cf. Table 2) represent the tribal classification according to Bremer and Manen

Table 2. Tribes included in or associated with the subfamily Rubioideae (Rubiaceae)

Tribes included in or associated with the Rubioideae by Bremekamp (1952, 1954, 1966), Verdcourt 1958, 1975), or Robbrecht (1988, 1993) and compared to the classification of Bremer and Manen. All investigated and accepted Rubioideae tribes are in boldface. Under the authors, tribal names are indicated with the first three letters of the tribal names; subfamilies are indicated with the first four letters of subfamilial names ANTIrheoideae, CINChonoideae, HILLioideae, POMAzotoideae, UROPhylloideae. "x": the tribe is accepted in the subfamily Rubioideae; "?": uncertain position according to the author; "in": the tribe is included in another tribe

	Bremekamp	Verdcourt	Robbrecht	Bremer and Manen
Tribes of Rubioideae				
Anthospermeae	X	X	X	X
Argostemmateae	X	X	X	X
Coussareeae	X	X	X	new cirumscription
Craterispermeae	ispermeae x x ANTI		ANTI	X
Danaideae				new tribe
Gaertnereae	X	in PSY?	in PSY	X
Lasiantheae				new tribe
Morindeae	X	X	X	X
Ophiorrhizeae	UROP	X	X	new circumscription
Paederieae	X	X	X	paraphyletic?
Psychotrieae	X	X	X	X
Rubieae	X	X	X	X
Schradereae	X	X	X	X
Spermacoceae	X	X	X	X
Theligoneae			X	X
Urophylleae	UROP	X	CINC	X
Included in other trib	es of Rubioideae			
Coccocypseleae	X	X	X	in COU
Cruckshanksieae	X	X	in HED	in COU
Hedyotideae	X	X	X	in SPE
Knoxieae	X	X	ANTI	in SPE
Manettieae	in HED	in HED	? in HED/CIN	in SPE
Opercularieae	?		in ANT	in ANT
Pauridiantheae	UROP	in URO	CINC	in URO
Pomazoteae	POMA	?	in HED	in OPH
Triainolepideae	X	in PSY	X	in SPE
Included in other Sub	families			
Hamelieae	X	X	X	CINC
Hillieae	HILL	?x	CINC	CINC
Virectarieae	in OPH	CINC	in HED	IXOR
Incertae sedis				
Lathraeocarpeae	X		X	? no material
Perameae	X		X	? no material

Inclusion based on morphology (cf. Puff and Igersheim 1994): *Metabolus* (incl. *Allaeophania*).

Tribe Coussareeae J. D. Hooker, in G. Bentham and J. D. Hooker, Genera Plantarum 2: 9, 24 (1873)

- Coccocypseleae Bremekamp, Rec. Trav.
 Bot. Neerl. 31: 253 (1934)
- Cruckshanksieae J. D. Hooker Genera Plantarum 2: 9, 20 (1873)

Useful recent studies: Dwyer (1966), Kirkbride (1976), Taylor (1996a)

Herbs (creeping in Coccocypselum), subshrubs, shrubs, or small trees. Stipules entire, cleft, as a ridge with appendages, or deeply bilobed. Flowers homo- or heterostylous, often 4-merous, white, blue, or bright yellow (Cruckshanksia, Oreopolus). Ovary 1-2-locular, with 1-2 or many (Coccocypselum) ovules per locule. Fruits often flattened (not Coccocypselum), fleshy, white (Coussarea) or often blue (Faramea, Coccocypselum) berries, (Declieuxia), schizocarps or thin-walled capsules (Cruckshanksia, Oreopolus), with 1-2 or many seeds, often flattened. Pollen 3-colporate or 2-4-porate. Chromosome basic number x = 10, 11? with 2- or 4-ploidy level (Kiehn 1995).

Genera included (here investigated): Coussarea, Coccocypselum, Declieuxia, Cruckshanksia, Faramea, Oreopolus.

In Andersson and Rova (1999) *Hindsia* and *Heterophyllaea* are nested within our Coussareeae. We have not had the possibility to sequence these genera, but *Hindsia* shows at least macromorphological similarity to *Declieuxia*. If this position is correct the fruit morphology of the tribe is even more variable as the fruits of *Hindsia* are capsular with hard walls and winged seeds.

Tribe Craterispermeae Verdcourt, Bull. Jard. Bot. État Brux. 28: 281 (1958)

Useful recent study: Igersheim (1992)

Shrubs or trees. Stipules entire. Inflorescences axillary and condensed and borne on a stout short peduncle. Flowers heterostylous. Ovary 2-locular, with a single pendulous ovule in each cell (one aborted?). Fruits fleshy (thickness of endocarp varies) with one seed. Pollen 3-colporate. Chromosome basic number x = 11 with 2-ploidy level (Kiehn 1995).

Genus included (here investigated): Craterispermum.

Tribe Gaertnereae Bremekamp ex S. Darwin, Taxon 25: 601 (1976)

Useful recent studies: Igersheim et al. (1994), Jansen et al. (1996)

Shrubs or trees. Stipules entire and united into a long cylindrical sheath, often with setae. Flowers often small and white. Ovary superior, 2-locular, with single erect ovule in each locule. Fruits fleshy, with one-seeded pyrenes. Pollen 2-3-colporate.

Genus included (here investigated): *Gaert-nera*.

Inclusion based on morphology (cf. Igersheim et al. 1994, and Jansen et al. 1996): *Pagamea*.

Tribe Morindeae Miquel, Flora van Nederlandsch Indie 2: 239, 241 (1857)

Useful recent study: Igersheim and Robbrecht (1993)

- Subtribe Morindinae de Candolle, Prodromus Systematis Naturalis 4: 342, 446 (1830)

Useful recent studies: Johansson (1988, 1994)

Shrubs, small trees, or lianas. Stipules entire or dentate, usually connate to sheathing. Flowers usually not heterostylous. Ovary 2-locular, with 2 erect ovules in each locule, later subdivided by secondary septum. Fruits drupaceous, often aggregated. Pollen 3-4-colporate. Chromosome basic number x = 11 with 2-, 4-, 8-, or 20-ploidy level (Kiehn 1995).

Genera included (here investigated): Morinda, Appunia, Coelospermum, Gynochthodes.

Inclusion based on morphology: Pogono-lobus.

Subtribe Prismatomerinae Ruang, Acta
 Phytotaxon. Sin. 26: 446 (1988)

Useful recent studies: Johansson (1987A, 1987B, 1989), Robbrecht et al. (1991)

Shrubs or trees. Stipules entire or often bilobed. Flower often heterostylous. Ovary 2-locular, with a single ovule in each locule. Fruits fleshy berries, often dark purple or blue, free or fused. Pollen 3-5-colporate.

Genus included (here investigated): Prismatomeris.

Inclusion based on morphology: Renellia (incl. Didymoecium), Motleyia.

- The Mitchella group

Useful recent study: Robbrecht et al. (1991)

Thorny shrubs or creeping herbs. Stipules entire or rarely bilobed. Flowers homo-(Damnacanthus) or hetero-stylous (Mitchella). Ovary 2-locular, with one pendulous ovule in each locule. Fruits fleshy, free or fused in pairs. Pollen 3-6-colporate. Chromosome basic number x = 11 with 2-ploidy level (Robbrecht, et al. 1991).

Genera included (here investigated): Mitchella, Damnacanthus.

Tribe Psychotrieae Cham. and Schltdl., Linnaea 4: 4. 1829

- Psathureae A. Rich. ex Dumort., Anal. Fam. Pl.: 32. 1829

Useful recent treatment: Taylor (1996b), Nepokroeff et al. (1999)

Shrubs, trees or herbs, or epiphytes. Stipules divided or rarely entire. Flowers often small, white and heterostylous. Ovary often 2-locular, with single erect ovule in each locule. Fruits fleshy, with one-seeded pyrenes. Seeds often with horny endosperm. Pollen very variable, 0-5-aperturate. Chromosome basic number x = (10), 11 with 2-12-ploidy level (Kiehn 1995).

Genera included (here investigated, an * indicates a paraphyletic taxon): Psychotria*, Amaracarpus, Cephaelis, Chasallia, Chazaliella, Geophila, Hydnophytum, Hymenocoleus, Myrmecodia, Palicourea*, Rudgea, Uragoga.

Inclusion based on morphology: Anthorrhiza, Myrmephytum, Squamellaria.

This tribe usually includes many genera (e.g. Robbrecht 1988) and *Psychotria* has been shown to be highly paraphyletic and many of the described genera are nested within it. Major revision of the whole tribe with new circumscriptions of genera will probably follow from initiated phylogenetic analysis (e.g. Nepokroeff pers. comm). Andersson and Rova (1999) have shown that *Readea*, *Streblosa* and *Margaretopsis* belong to this group of taxa but we have not sequenced these genera and it is not obvious from the morphology that they belong here.

Tribe Schradereae Bremekamp, Rec. Trav. Bot. Neerl. 31: 253 (1934)

Useful recent studies: Puff et al. (1993), Puff and Buchner (1998), Puff et al. (1998a), Puff et al. (1998b)

Shrub or often epiphytic climbers with adhesive roots. Stipules entire. Ovary 2-(3-4)-locular. Flowers in congested spherical inflorescences, heterostyly has been reported. Fruits fleshy, baccate, with many seeds. Chromosome basic number x = 11 with 2-ploidy level (Kiehn 1995). Pollen 2-3-(4-)-porate.

Genera included (here investigated): Schradera.

Inclusion based on morphology (cf. Puff et al. 1993): Lecananthus, Leucocodon.

Tribe Danaideae B. Bremer and Manen, trib. nov.

Type genus: Danais

Useful recent studies: Buchner and Puff (1993), Puff and Buchner (1994)

Lianae lignosae, frutices interdum scandentes vel arbores parvae. Stipulae integrae, bifidae vel fimbriatae. Flores heterostyli. Corollae valvatae vel reduplicatae-valvatae. Ovaria 2-locularia in quoque loculo ovulis numerosis. Fructus capsulares loculicidi vel septicidales seminibus alatis. Pollen 3-4 (-5)-colporatum. — Differt a tribuluis ceteris habitibus plerumque scandentibus lignosis et seminibus alatis (itidem in *Bouvadria* et *Manettia*; Spermacoceae).

Woody lianas, climbing shrubs, shrubs, or small trees. Flowers heterostylous. Stipules entire, bifid or fimbriate. Corolla valvate or valvate-reduplicate (*Danais* and *Schismatoclada*). Ovary 2-locular with numerous ovules in each locule. Fruits capsular loculicidal (*Danais* and *Payera*) or septicidal (*Schismatoclada*), with winged seeds. Pollen 3-4(-5)-colporate.

Genus included (here investigated): *Danais*. Inclusion based on morphology: (cf. Buchner and Puff 1993) *Schismatoclada* (inclusion supported by unpublished *rbc*L data: Bremer), *Payera*.

Tribe Spermacoceae Bercht. and J. Presl, Prir. Rostlin: 256 (1820) – the change of the authors from earlier A. Rich. ex Dumort., Analyse des familles des plantes: 33 (1829) was first noted by J. Reveal (pers. comm.)

- Hedyotideae Chamisso and Schlechtendal ex de Candolle, Prodromus Systematis Naturalis 4: 342, 401(1830)
- Knoxieae Hooker f. in Bentham and Hooker, Genera plantarum 2: 9, 21 (1873)
- Manettieae Bremekamp, Rec. Trav. Bot.
 Neerl. 31: 253 (1934)
- Triainolepideae Bremekamp, Proc. Kon. Akad. Wetensch. 59: 3 (1956)

Useful recent studies: Puff (1988), Puff and Robbrecht (1989), Mena (1990)

Herbs or subshrubs (a few species are small trees). Stipules fimbriate. Flowers quite often heterostylous. Ovary 1-5- or often 2-locular, with many to single ovules in each locule. Fruits dry, dehiscent or indehishent, or rarely fleshy (e.g. *Triainolepis*) with many to single seeds. Pollen often 3-colporate-pluricolpate. Chromosome basic number very variable x = 6-17 with 2-20-ploidy levels (Kiehn 1995).

Genera included (here investigated, an * indicates a paraphyletic taxon): Spermacoce* (incl. Borreria), Agathisanthemum, Amphiasma, Arcytophyllum, Batopedina Bouvardia, Carphalea (incl. Dirichletia), Conostomium, Diodia, Ernodea, Hedyotis* (incl.

Kadua), Hemidiodia, Knoxia, Kohautia, Lelya, Manettia, Mitracarpus, Oldenlandia*, Otiophora, Otomeria, Parapentas, Paratriaina, Pentanisia, Pentanopsis, Pentas, Pentodon, Phylohydrax, Placopoda, Psyllocarpus, Richardia, Triainolepis.

A majority of the former Hedyotideae, Knoxieae, and Spermacoceae s.s. genera should be included in this tribe. Inclusion based on morphology is supported for at least the following genera: Chaemepentas, Crusea, Dentella, Dibrachionostylus, Dolichometra, Hedythyrsus, Hydrophylax, Lucya, Manostachya, Mitrasacmopsis, Neohymenopogon, Nodocapaea, Pseudohedyotis, Schwendera, Synaptantha, Staelia, Thecorchus, Thyridocalyx.

Tribe Anthospermeae Chamisso and Schlechtendal ex de Candolle, Prodromus Systematis Naturalis 4: 343, 578 (1830)

- Opercularieae A. Rich. ex de Candolle,
 Prodromus Systematis Naturalis 4: 343, 614 (1830)
- Durringtonieae Henderson and Guymer, Kew Bull. 40: 97–107 (1985)

Useful recent studies: Puff (1982, 1986), Robbrecht (1982)

Herbs, shrubs, dwarf shrubs, or small trees. Stipules entire or divided. Flowers usually unisexual, anemophilous, not heterostylous. Stamens usually inserted low in the corolla. Stigma long and filiform. Ovary 1-2-(5-)locular, with a single erect ovule in each locule. Fruits dry and splitting into cocci or capsular, or fleshy. Chromosome basic number x = 11 (one exception of 10) with 2-20-ploidy levels (Kiehn 1995). Pollen 3-colporate.

Genera included (here investigated): Anthospermum, Carpacoce, Coprosma, Galopina, Nenax, Nertera, Opercularia, Phyllis.

Inclusion based on morphology: Durringtonia, Eleutheranthus, Leptostigma (incl. Corynula), Normandia, Peratanthe, Pomax (inclusion of Pomax supported by unpublished rbcL data Bremer).

Tribe Argostemmateae Bremekamp ex Verdcourt, Bull. Jard. Bot. État Brux. 28: 281 (1958)

Useful recent study: Bremer (1989)

Herbs with iso- or anisophyllous leaves. Stipules entire or slightly cleft. Flowers hermaphroditic. Stamens usually inserted at base of the corolla, adnate into an anther cone or free. Anthers open with vertical slits or rarely with pores. Ovary 2-6-locular. Fruit a succulent capsule opening by an apical operculum (Argostemma) or a berry (Mycetia), with many small seeds. Chromosome basic number x = 11, 14 with 2- or 4-ploidy levels (Kiehn 1995).

Genera included (here investigated): Argostemma, Mycetia.

Tribe Paederieae de Candolle, Prodromus Systematis Naturalis 4: 342, 470 (1830)

- Putorieae de Candolle ex Sweet, Sweet's Hortus Britannicus, ed 3, 325 (1839)

Useful recent studies: Puff (1982), Thulin (1997)

Shrubs, climbers, dwarf shrubs and herbs, many with a foetid smell. Stipules divided or entire. Flowers often heterostylous, Stamens inserted in upper part of corolla, Ovary 2-5-locular, with a single erect ovule in each locule. Fruits dry, dehishent into mericarps or opening with operculum, or fleshy, indehiscent. Chromosome basic number x = 11 (rarely 10, 12, 13) with 2-, 4-, 6- or 8-ploidy levels (Kiehn 1995). Pollen 3-colpate.

Genera included (here investigated): Paederia, Gaillonia, Plocama, Putoria, Serissa, Spermadictyon.

Inclusion based on morphology: Choulettia, Jaubertia, Kelloggia, Leptodermis, Pseudogaillonia, Pseudopyxis, Pterogaillonia.

Tribe Theligoneae Wunderlich ex S. Darwin, Taxon 25: 607 (1976)

Useful recent study: Wunderlich (1971)

Herbs with upper leaves alternate, anisophyllous. Stipules sheathing. Flowers unisexual, anemophilous. Male flowers usually with many, up to 30 stamens. Ovary 1-locular, with

one ovule. Fruits fleshy, with elaiosome and a single seed. Pollen (3-)4-8-porate. Chromosome basic number x = 10, 11 with 2-ploidy level (Kiehn 1995).

Genus included (here investigated): *Theligonum*.

Tribe Rubieae Baill., Hist. Pl. 7: 365, 390 (1880)

- Galieae A. Rich. ex Dumort., Anal. Fam. Pl.: 33. (1829)
- Asperuleae A. Rich., Mém. Soc. Hist. Nat. Paris 5: 126 (1830)

Useful recent studies: Manen et al. (1994), Natali et al. (1996)

Herbs or rarely subshrubs, with polygonal stems. Leaves (and or leaf-like stipules) verticillate. Flowers generally perfect and heterostylous, *Phuopsis* with secondary pollen presentation. Calyx rudimentary. Ovary 2-locular, with single ovule in each locule. Fruits dry or fleshy, didymous, seeds usually adhering to the pericarp. Pollen pluricolpate. Chromosome basic number x = 9-12 with 2-12-ploidy levels.

Genera included (here investigated, an * indicates a paraphyletic taxon): Rubia, Asperula*, Galium*, Relbunium, Didymaea, Cruciata, Phuopsis, Sherardia, Valantia.

Inclusion based on morphology: Callipeltis, Crucianella, Mericarpaea (inclusion supported by unpublished atpB-rbcL data: Manen), Microphysa, Warburgia.

Discussion

The power of the present study is that the phylogeny and proposed classification is based on many characters from different molecular markers. One is protein-coding (*rbc*L), one is a non-coding spacer (the *atp*B-*rbc*L spacer), and the third is a non-coding intron marker (*rps*16 from Andersson and Rova 1999).

Another strength of our study is the comprehensive tribal sampling, e.g., we include, for the first time, representatives of the tribe Craterispermeae and the former tribes Pomazoteae and Triainolepideae. Our analysed taxa represent all except two of the 28

Rubiaceae tribes that have been associated with the subfamily Rubioideae. We accept 16 in Rubioideae (Table 2), nine are included as synonyms, three are included in other subfamilies and two have been left "incertae sedis" as we did not manage to get any material. If compared to earlier classifications of the subfamily (Bremekamp 1954, 1966; Verdcourt 1958; Robbrecht 1988, 1993) it seems that Verdcourt's scheme is best supported by our data and our classification is in many parts similar to his. Of our 16 tribes 12 more or less correspond to earlier tribal circumscriptions, two tribes have received new very different circumscriptions, and two are new to science.

The topologies of the trees are almost identical or congruent in our four analyses. The resolution and support are highest in the combined analyses, which is expected as more characters are included (cf. Bremer et al. 1999). In all analyses (Figs. 1-4) the same seven tribes are monophyletic: Argostemmateae, Coussareeae, Ophiorrhizeae (3-data matrix includes only one taxon), Psychotrieae, Rubieae, Spermacoceae, and Urophylleae. Furthermore, Anthospermeae have good support in the rbcL (Fig. 2) and combined analyses (Fig. 3) but not in the atpB-rbcL data (Fig. 1). Lasiantheae are represented by two taxa in the rbcL analysis (but only one in the other analyses) and are highly supported (100%; Fig. 2). Five tribes are represented by single taxa or are monogeneric and thus the monophyly can not be tested: Craterispermeae, Danaideae, Gaertnereae, Schradereae, and Theligoneae. Morindeae are monophyletic in the combined analyses (but not supported <50%; Figs. 3, 4). Finally, the tribe Paederieae is not supported in our data, but instead paraphyletic. However, at this moment, we have taken a conservative approach and maintain Paederieae as a tribe while awaiting a more detailed study (under revision by M. Backlund pers. comm.).

Our results are also in many parts similar to those from the recent study by Andersson and Rova (1999) of *rps*16 data, but, there are important differences. Notably, the results

from our combined analysis including their rps16 data are different from their results. However, in our analysis the rps16 data are a minor part of the matrix. Other explanations might be that we use different alignments, or different ways of analysis. They never completed their analysis as they ran out of computer memory when 14 600 trees were found. Had it been possible to save all equally parsimonious trees, their consensus could be much more collapsed, so it is difficult to judge to what extent the rps16 data alone are phylogenetically informative. Clades not supported by bootstrap values well over 50% (preferably 75%) in the trees presented by Andersson and Rova should not be considered reliable. Two examples concern the circumscription and position of Spermacoceae and Theligoneae, respectively.

Andersson and Rova (1999) found what we call tribe Spermacoceae paraphyletic. The incongruence between our studies must be of the "soft incongruency" type (Seelanaen et al. 1997). Our tribe Spermacoceae is monophyletic and supported in most of our analyses (in atpB-rbcL not supported but congruent, rbcL 99%, rbcL/atpB-rbcL 99%, rbcL/atpB-rbcL/ rps16 100%). Andersson and Rova illustrate (in one out of 14 600 trees) the Knoxieae (in our data included in the Spermacoceae) as the sister group to the rest of the Spermacoceae alliance. However, this relationship is not supported by their data (<50%). Andersson and Rova also found Theligoneae sister to Plocama, a member of the Paederieae (56%). In our analyses the sister group relationship of Theligoneae is not *Plocama* but with the Rubieae, strongly supported by our data (rbcL 88%, rbcL/atpB-rbcL 96%, rbcL/atpB-rbcL/ rps16 97%, not supported by the atpB-rbcL). This difference is of the "hard incongruency" type (Seelanaen et al. 1997).

The following discussion of phylogeny and classification is based on the tree from the combined analysis of the *rbcL/atpB-rbcL* data (Fig. 3), unless otherwise stated, since this analysis has the best taxon sampling relevant for the tribal phylogeny.

The basal clades. The first dichotomy of the ingroup (Fig. 3) in the tree is between a newly circumscribed Ophiorrhizeae and the rest of the subfamily. In agreement with Bremer (1996a), the genus Neurocalyx (formerly Argostemmateae cf. Bremer 1987) belongs to this clade and for the first time we here show that the genera Lerchea and Xanthophytum formerly in Pomazoteae (Bremekamp 1966) or Hedyotideae (Robbrecht 1993) also are closely related. The tribe is supported by a high bootstrap value (92%) and is distinctly separated (87%) from the rest of the subfamily. One uniting morphological character for the taxa of the Ophiorrhizeae is the occurrence of very small seeds (dust seeds). Andersson and Rova (1999) found Ophiorrhiza to be sister to Urophylleae and mention that the subfamily Urophylloideae (incl. Ophiorrhiza) "is defensible, being the sister group to the rest of the Rubioideae, but it may not be a useful entity". We certainly agree with the latter statement but disagree with their recognition of the subfamily Urophylloideae. Our data do not support inclusion of *Ophiorrhiza* in the same clade as the Urophylleae. The differences in results might be due to taxon sampling and analysis methods. We included more representatives of Ophiorrhizeae (4 genera compared to 1) and Urophylleae (5 genera compared to 3) and our analyses run to completion. Morphologically Ophiorrhizeae differ from Urophylleae by type of fruits and habit, Urophylleae are woody with fleshy berries while Ophiorrhizeae never have berries and are often herbaceous.

The next branch to split off (100% bootstrap value) is the tribe Urophylleae, with Pauridiantha nested within the tribe (as in Andersson and Rova 1999). Inclusion of Pauridiantha within Urophylleae was first proposed by Verdcourt (1958) in contrast to Bremekamp (1954, 1966) and Robbrecht (1988, 1993), who recognised Pauridiantheae as a separate tribe next to the Urophylleae. These two tribes were included in the separate subfamily Urophylloideae by Bremekamp. Robbrecht did not accept this subfamily and

he moved Urophylleae and Pauridiantheae to subfamily Cinchonoideae. Verdcourt was the first author to include the Urophylleae in the Rubioideae which is in concordance with the present study. The Urophylleae are characterised by berries with many seeds.

Lasianthus is the next branch to split off. It is distinctly separated from the Urophylleae part of the tree (97%) and the rest of the subfamily (100%). The genus was earlier included in Psychotrieae (by most earlier authors) or in Morindeae (Robbrecht 1988, 1993). A position within the Psychotrieae alliance has earlier been refuted by Bremer (1996a). As Lasianthus in our study is the sister to the rest of the subfamily (confirmed by an unpublished rbcL sequence of Lasianthus strigosus, Bremer) and unless all the remaining Rubioideae taxa should be lumped into one tribe, Lasianthus must form a new tribe. Further, the genus Trichostachys belongs to this distinct branch (rbcL analysis). The Lasiantheae are characterised by fleshy drupes with 2 to 12 pyrenes, often blue or black. The position of Lasiantheae is supported by the 3data matrix and by the results of Andersson and Rova (1999). In their analysis the Lasianthus branch is sister to Perama of the tribe Perameae (supported by 100%). If this relationship is correct one could argue that Lasianthus should be included in the tribe Perameae instead of being elevated to a new tribe Lasiantheae. However, we agree with Andersson and Rova (1999) that such a taxon would be morphologically undefinable. Perama is a genus of tiny, hairy plants, with very small or reduced stipules, flowers in terminal heads, calyx of two lobes, and dry 3-locular capsular fruits with a single ovule in each locule. The single ovule is the only similarity to Lasiantheae, but solitary ovules occur and have evolved several times in the subfamily.

The next node includes the Coussareeae (100%) as sister to the Psychotrieae alliance (99%) and the Spermacoceae alliance (100%), a relationship with low support (54%). For the Coussareeae a new, highly unexpected, relationship was shown between *Faramea* (Cous-

sareeae) and Coccocypselum (former Coccocypseleae) by Bremer (1996a, Coussarea was also added to this clade in Bremer and Thulin 1998). Further, Nepokroeff et al. (1999) and Andersson and Rova (1999) showed that the genus Declieuxia, formerly in Psychotrieae also belongs to this group. Another interesting result from the present study (and Andersson and Rova 1999) is the position of Cruckshanksia (rbcL analysis) and Oreopolus (rbcL and atpB-rbcL analyses) also placed in this same clade. These last two genera have been accepted as a separate tribe Cruckshanksieae or they have been included in Hedvotideae (cf. Table 2). We have decided to include all these six genera in the tribe Coussareeae despite morphological variation (large parts of the variation is found within the "Coussareeae s.s." and the "Coccocypseleae" branches, respectively). There are morphological traits that support their relationship, e.g., often 4merous flowers, flattened berry-like or thinwalled capsular fruits, and flattened seeds. Earlier Coussareeae were supposed to have drupaceous fruits which has been refuted (C. Taylor pers. comm.). Interestingly all Coussareeae genera are American and most of them tropical. The African genus Schizocolea was earlier included in Coussareeae but unpublished rbcL data (Bremer) contradict that position.

The Psychotrieae alliance is well-supported with a high bootstrap value (99%; Fig. 3). This group comprises many species (probably ca 2000) which can be classified in five tribes: Psychotrieae, Craterispermeae, Gaertnereae, Morindeae, and Schradereae. They are characterised by fleshy drupes with one ovule per carpel (one or two in Morindeae). Many are important as food sources for frugivorous birds in the tropics (cf. Snow 1981). The relationships and delimitations of the *Psycho*tria alliance are still unclear and need further investigations. The largest tribe is the Psychotrieae s.s. which is distinctly separated from the other taxa (83% bootstrap values). Molecular data (Bremer 1996a) have earlier shown that the type genus of the tribe, Psychotria, is paraphyletic. It has recently been demonstrated by Nepokroeff et al. (1999) that *Psychotria* together with the other genera of the tribe nicely can be split into minor monophyletic groups, e.g., one includes *Psychotria* s.s. (subgenera *Psychotria* and *Tetramerae* and a Pacific group including e.g. the myrmecophilous genera *Anthorrhiza*, *Hydnophytum*, *Myrmecodia*, and *Myrmephytum*), another group includes the subgenus *Heteropsychotria* and the genus *Palicourea*. Also other genera such as *Chasallia*, *Chazaliella*, *Geophila*, *Rudgea*, and *Hymenocoleus* belong to this tribe.

Since the relationships and circumscriptions of the other tribes of the Psychotria alliance group are uncertain it is premature to propose a classification. However, from our results it is evident that the tribes Craterispermeae, Gaertnereae, Schradereae, and Morindeae all belong to this group and that they are distinctly separated from Psychotrieae. Further, all these tribes have been described before, are accepted by several authors, and are morphologically distinct. We find it more informative to accept them as tribes than sink them in Psychotrieae. Craterispermeae were placed close to Psychotrieae by Verdcourt (1958) but Robbrecht removed them to subfamily Antirheoideae, a position not supported by our data. Further, from the present study as well as from Nepokroeff et al. (1999) and Andersson and Rova (1999) there are no indications that the genus Gaertnera should be included in the tribe Psychotrieae as was suggested e.g. by Verdcourt (1958, although with some hesitation) and Robbrecht (1988, 1993). The Schradereae have recently been expanded to include also Lecananthus, Leucocodon (Puff and Buchner 1998, Puff et al. 1998, Puff et al. 1998) and it seems correct that they form a distinct monophyletic group. A monophyletic tribe Morindeae is not contradicted by our data but there is no support for the tribe either (<50%). The delimitation of and relationships within the Morindeae have been subject to recent discussions (e.g. Igersheim and Robbrecht 1993). In our study we find the Morindeae representatives in three groups, corresponding to subtribe Morindinae, subtribe Prismatomerinae, and the Mitchella group (Robbrecht 1993). The relationship between Mitchella and Damnacanthus of the Mitchella group was first pointed out by Robbrecht et al. (1991) and was also highly supported in Bremer (1996a). Andersson and Rova (1999) included four of the tribes in the Psychotrieae alliance but no representative of Craterispermeae. Their sample tree is fully resolved. Psychotrieae and Gaertnereae are shown to be well-supported, but the relationships between the Gaertnereae, Schradereae, and Morindeae have only low support (59%, 55%). The monophyly of Morindeae has good support (80%), but Andersson and Rova did not include any representative of the subtribe Prismatomerinae. In our 3-data analysis the support for the Morindeae (including Prismatomeridae is very low (48%).

The Spermacoceae alliance includes the rest of the subfamily Rubioideae. It is a strongly supported clade (100%; Fig. 3) and represents a majority of the dry-fruited taxa of the subfamily. In this group we accept seven tribes and of these six (Spermacoceae, Anthospermeae, Argostemmateae, Danaideae, Rubieae, Theligoneae) are distinctly separated from each other with bootstrap values ranging from 99% to 100%. One tribe, the Paederieae, is found to be biphyletic. As the tribe is under revision (M. Backlund pers. comm.) we refrain from doing any taxonomic changes at the moment.

At the most basal dichotomy in this Spermacoceae alliance we find the genus *Danais*. It is one of the genera with winged seeds that Bremekamp (1952) transferred from the Cinchoneae to the Hedyotideae (here synonym to Spermacoceae), a position that was accepted by Verdcourt (1958). Robbrecht (1988, 1993) on the other hand considered it a link between Rubioideae and Cinchonoideae. In a detailed study of *Danais* Buchner and Puff (1993) showed it to be closely related to *Schismatoclada* and *Payera*. The close relationship of *Danais* and *Schismatoclada* has been confirmed by sequence data (Bremer unpublished). The present analysis supports the conclusion from

Bremer (1996a) that *Danais* belongs to the Rubioideae, and is nested within the subfamily as the sister group to the rest of the Spermacoceae alliance separated from the other tribes and as there are certain morphological characters that are rather rare in this part of Rubiaceae (woody lianas, climbing shrubs or shrubs to small trees with capsular fruits containing many winged seeds), we do not hesitate to propose a new tribe.

The next node in the tree is a split between the tribe Spermacoceae (fide Bremer 1996a) and the rest of the tribes. The Spermacoceae includes all investigated taxa of the former tribes Hedyotideae (excluding Danais as noted above), Manettieae, Knoxieae, and Spermacoceae and further also Triainolepis of the former tribe Triainolepideae. None of these tribes can be accepted as often circumscribed (Verdcourt 1958; Bremekamp 1966; Robbrecht 1998, 1993). Within this large group of taxa there is a basal split into two highly supported branches (100% bootstrap fractions for each), one representing the Pentas/Triainolepis group and the other including the rest of the clade, here called the Hedyotis/Spermacoce group. These two groups were initially identified by morphological characters (Bremer 1987) and supported by molecular data (Bremer et al. 1995, Bremer 1996a). The Pentas/Triainolepis group includes also the tribe Knoxieae (represented by *Pentanisia*), a tribe Robbrecht (1988) transferred to the subfamily Antirheoideae. Our placement of Pentanisia, and the former Knoxieae, is corroborated by the rbcL analysis which also includes *Knoxia* in the same branch. One of the characters for Knoxieae was solitary pendulous ovules, but at least ovule reduction occurs several times in this part of the Rubioideae, also in the closely related Placopoda and Carphalea. In the Hedyotis/ Spermacoce group many of the genera have numerous ovules and wing-less seeds, e.g., Hedyotis, Oldenlandia, Kohautia, and Pentodon, but there are also genera with many ovules and winged seeds, earlier included in Manettieae (Bouvardia and Manettia) as well as the genera of the tribe Spermacoceae s.s.

with solitary, erect ovules. As shown before (Bremer 1996a), the large genera *Hedyotis* and *Oldenlandia* are paraphyletic. The position of Spermacoceae s.s. within the former Hedyotideae (pointed out by Bremer et al. 1995 and Natali et al. 1995, Bremer 1996a) made the Hedyotideae paraphyletic.

The next clade in the tree corresponds to the tribe Anthospermeae, supported in the combined analyses (100% in both analyses), the *rbc*L analysis (74%), and in Andersson and Rova (1999; 54%). However, in our *atp*B-*rbc*L tree there is no support for the tribe.

The next branch represents the new circumscription of Argostemmateae (fide Bremer 1996a), well-supported (100%), so also in Andersson and Rova (1999; 99%). The former circumscription of the tribe (including Neurocalyx and Steenisia) was mainly based on a supposedly unique character, adnate anthers. However, that character is homoplastic and has evolved at least three times in the family (in Argostemma, Neurocalyx of the Ophiorrhizeae, and Steenisia in the subfamily Cinchonoideae, Bremer 1984).

Sister to Argostemmateae is a clade containing the tribes Rubieae, Theligoneae, and representatives of the tribe Paederieae. The circumscription and taxonomic position of the tribe Paederieae has been much discussed, e.g., Puff (1982) was of the opinion that the tribe is close to Theligoneae and Anthospermeae and he transferred all insect-pollinated genera from Anthospermeae to Paederieae. Today the tribe includes ca. 15 genera (Robbrecht 1988, 1993) but it is difficult to identify characters that unite all taxa, as most characters are variable. We have investigated five genera molecularly (Gaillonia, Plocama, Putoria, Serissa, Spermadictyon). The analyses indicate that the taxa

are related to the Rubieae and Theligoneae but not to Anthospermeae, nor do they form a natural group; instead they constitute a grade (same result in Andersson and Roya 1999).

The monogeneric tribe Theligoneae is the sister group to the Rubieae. This close relationship was earlier indicated by Bremer et al. (1995) and Natali et al. (1995), although contradicted in Andersson and Rova (1999), however, with low support. Finally, the Rubieae is supported by 100% in all our analyses. The strong support for this group is congruent with earlier results (Natali et al. 1995, Bremer 1996a, Andersson and Rova 1999).

Representatives of the Lathraeocarpeae remain to be investigated and the recently proposed position of the Perameae (Andersson and Rova 1999) needs to be confirmed. Material of these remaining taxa has so far been difficult to obtain or analyse. With this study we have nevertheless provided a comprehensive phylogeny of Rubioideae and most of the tribes earlier associated with the subfamily have been investigated. A classification including ca 150 genera in 16 well-supported monophyletic tribes summarises our results.

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Appendix 1. Genera included in the Rubioideae, fide Bremer and Manen. A list of genera sorted within tribes in alphabetical order. Tribal position is indicated with the first three letters of the tribal name. Morindeae (MOR) contain also: -M, -M1 or -P indicating subgroups. *paraphyletic taxon

Genus	Tribe	Genus	Tribe	Genus	Tribe
Anthospermum	ANT	Kelloggia	PAE	Dibrachionostylus	SPE
Carpacoce	ANT	Plocama	PAE	Diodia	SPE
Coprosma	ANT	Pseudogaillonia	PAE	Dirichletia	SPE
Durringtonia	ANT	Pterogaillonia	PAE	Dolichometra	SPE
Eleutheranthus	ANT	Putoria	PAE	Ernodea	SPE
Galopina	ANT	Leptodermis	PAE	Hedyotis*	SPE
Leptostigma	ANT	Pseudopyxis	PAE	Hedythyrsus	SPE
Nenax	ANT	Serissa	PAE	Hemidiodia	SPE
Nertera	ANT	Spermadictyon	PAE	Houstonia	SPE
Normandia	ANT	Âmaracarpus	PSY	Hydrophylax	SPE
Opercularia	ANT	Anthorrhiza	PSY	Kadua	SPE
Peratanthe	ANT	Cephaelis	PSY	Knoxia	SPE
Phyllis	ANT	Cĥasallia	PSY	Kohautia	SPE
Pomax	ANT	Chazaliella	PSY	Lelya	SPE
Argostemma	ARG	Geophila	PSY	Lucya	SPE
Mycetia	ARG	Hydnophytum	PSY	Manettia	SPE
Coccocypselum	COU	Hymenocoleus	PSY	Manostachya	SPE
Coussarea	COU	Myrmecodia	PSY	Mitracarpus	SPE
Cruckshanksa	COU	Myrmephytum	PSY	Mitrasacmopsis	SPE
Declieuxia	COU	Palicouria	PSY	Neohymenopogon	SPE
Faramea	COU	Psychotria*	PSY	Nodocapaea	SPE
Oreopolus	COU	Rudgea	PSY	Oldenlandia*	SPE
Craterispermum	CRA	Squamellaria	PSY	Otiophora	SPE
Danais	DAN	Uragoga	PSY	Otomeria	SPE
Payera	DAN	Asperula*	RUB	Parapentas	SPE
Schismatoclada -	DAN	Callipeltis	RUB	Paratriaina	SPE
Gaertnera	GAE	Crucianella	RUB	Pentanisia	SPE
Pagamea	GAE	Cruciata	RUB	Pentanopsis	SPE
Lasianthus	LAS	Didymaea	RUB	Pentas	SPE
Trichostachys	LAS	Galium*	RUB	Pentodon	SPE
Metabolus	LAS	Mericarpaea	RUB	Phylohydrax	SPE
Appunia	MOR-M	Microphysa	RUB	Placopoda	SPE
Coelospermum	MOR-M	Phuopsis	RUB	Pseudohedyotis	SPE
Gynochthodes	MOR-M	Relbunium	RUB	Psyllocarpus	SPE
Morinda	MOR-M	Rubia	RUB	Richardia	SPE
Pogonolobus	MOR-M	Sherardia	RUB	Schwendera	SPE
Damnacanthus	MOR-MI	Valantia	RUB	Spermacoce*	SPE
Mitchella	MOR-MI	Warburgia	RUB	Staelia	SPE
Motleyia	MOR-P	Lecananthus	SCH	Synaptantha	SPE
Prismatomeris	MOR-P	Leucocodon	SCH	Thecorchus	SPE
Renellia	MOR-P	Schradera	SCH	Thyridocalyx	SPE
Coptophyllum	OPH	Agathisanthemum	SPE	Triainolepis	SPE
Lerchea	OPH	Amphiasma	SPE	Theligonum	THE
Neurocalyx	OPH	Arcytophyllum	SPE	Amphidasya	URO
Ophiorrhiza	OPH	Batopedina	SPE	Commitheca	URO
Spiradiclis	OPH	Bouvardia	SPE	Maschalocorymus	URO

Appendix 1. (Continued)

Genus	Tribe	Genus	Tribe	Genus	Tribe
Xanthophytum Paederia Choulettia Gaillonia Jaubertia	OPH PAE PAE PAE PAE	Carphalea Chaemepentas Conostomium Crusea Dentella	SPE SPE SPE SPE SPE	Pauridiantha Praravinia Pravinaria Urophyllum	URO URO URO URO

Appendix 2. Genera included in the Rubioideae, fide Bremer and Manen. A list of genera in alphabetical order. Tribal position is indicated with the first three letters of the tribal name. Morindeae (MOR) contain also: -M, -M1 or -P indicating subgroups. *paraphyletic taxon

Genus	Tribe	Genus	Tribe	Genus	Tribe
Agathisanthemum	SPE	Hedyotis*	SPE	Pauridiantha	URO
Amaracarpus	PSY	Hedythyrsus	SPE	Payera	DAN
Amphiasma	SPE	Hemidiodia	SPE	Pentanisia	SPE
Amphidasya	URO	Houstonia	SPE	Pentanopsis	SPE
Anthorrhiza	PSY	Hydnophytum	PSY	Pentas	SPE
Anthospermum	ANT	Hydrophylax	SPE	Pentodon	SPE
Appunia	MOR-M	Hymenocoleus	PSY	Peratanthe .	ANT
Arcytophyllum	SPE	Jaubertia	PAE	Phuopsis	RUB
Argostemma	ARG	Kadua	SPE	Phyllis	ANT
Asperula*	RUB	Kelloggia	PAE	Phylohydrax	SPE
Batopedina	SPE	Knoxia	SPE	Placopoda	SPE
Bouvardia	SPE	Kohautia	SPE	Plocama	PAE
Callipeltis	RUB	Lasianthus	LAS	Pogonolobus	MOR-M
Carĥpalea	SPE	Lecananthus	SCH	Pomax	ANT
Carpacoce	ANT	Lelya	SPE	Praravinia	URO
Cephaelis	PSY	Leptodermis	PAE	Pravinaria	URO
Chaemepentas	SPE	Leptostigma	ANT	Prismatomeris	MOR-P
Chasallia	PSY	Lerchea	OPH	Pseudogaillonia	PAE
Chazaliella	PSY	Leucocodon	SCH	Pseudohedyotis	SPE
Choulettia	PAE	Lucya	SPE	Pseudopyxis	PAE
Coccocypselum	COU	Manettia	SPE	Psychotria*	PSY
Coelospermum	MOR-M	Manostachya	SPE	Psyllocarpus	SPE
Commitheca	URO	Maschalocorymus	URO	Pterogaillonia	PAE
Conostomium	SPE	Mericarpaea	RUB	Putoria	PAE
Coprosma	ANT	Metabolus	LAS	Relbunium	RUB
Coptophyllum	OPH	Microphysa	RUB	Renellia	MOR-P
Coussarea	COU	Mitchella	MOR-MI	Richardia	SPE
Craterispemum	CRA	Mitracarpus	SPE	Rubia	RUB
Cruckshanksieae	COU	Mitrasacmopsis	SPE	Rudgea	PSY
Crusea	SPE	Morinda	MOR-M	Schismatoclada	DAN
Crucianella	RUB	Motleyia	MOR-P	Schradera	SCH
Cruciata	RUB	Mycetia	ARG	Schwendera	SPE
Damnacanthus	MOR-MI	Myrmecodia	PSY	Serissa	PAE
Danais	DAN	Myrmephytum	PSY	Sherardia	RUB
Declieuxia	COU	Nenax	ANT	Spermacoce*	SPE
Dentella	SPE	Neohymenopogon	SPE	Spermadictyon	PAE

Appendix 2. (Continued)
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Genus	Tribe	Genus	Tribe	Genus	Tribe
Dibrachionostylus	SPE	Nertera	ANT	Spiradichlis	OPH
Didymaea	RUB	Neurocalyx	OPH	Squamellaria	PSY
Diodia	SPE	Nodocapaea	SPE	Staelia	SPE
Dirichletia	SPE	Normandia	ANT	Synaptantha	SPE
Dolichometra	SPE	Oldenlandia*	SPE	Thecorchus	SPE
Durringtonia	ANT	Opercularia	ANT	Theligonum	THE
Eleutheranthus	ANT	Ophiorrhiza	OPH	Thyridocalyx	SPE
Ernodea	SPE	Oreopolus	COU	Triainolepis	SPE
Faramea	COU	Otiophora	SPE	Trichostachys	LAS
Gaertnera	GAE	Otomeria	SPE	Uragoga	PSY
Gaillonia	PAE	Paederia	PAE	Urophyllum	URO
Galium*	RUB	Pagamea	GAE	Valantia	RUB
Galopina	ANT	Palicouria	PSY	Warburgia	RUB
Geophila	PSY	Parapentas	SPE	Xanthophytum	OPH
Gynochthodes	MOR-M	Paratriaina	SPE		

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